Rat Luteinizing Hormone ELISA Kit

Cat #: orb1473274 (manual)

Competitive Inhibition Enzyme Immunoassay for quantitative detection of Rat Luteinizing Hormone concentrations in Serum, Plasma or Other Biological Fluids.

For research use only. Not for diagnostic or therapeutic procedures.

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INTRODUCTION

Luteinizing hormone (LH) is produced in both men and women from the anterior pituitary gland in response to luteinizing hormone-releasing hormone (LH-RH or Gn-RH), that is released by the hypothalamus. LH, also called interstitial cell-stimulating hormone (ICSH) in men, is glycoprotein with a molecular weight of approximately 30,000 Dalton. LH stimulates ovulation and ovarian steroid production in the female. In the male, LH controls Leydig cell secretion of testosterone. LH is elevated in Luteal phase of menstrual cycle, primary hypogonadism, Gonadotropin-secreting pituitary tumors and menopause. LH is deceased in hypothalamic Gn-RH deficiency, pituitary LH deficiency and ectopic steroid production.

ASSAY PRINCIPLES

Biorbyt Rat Luteinizing Hormone ELISA Kit (Competitive Inhibition Enzyme Immunoassay) is a competitive inhibition enzyme immunoassay technique for the in vitro quantitative measurement of Rat Luteinizing Hormone in Serum, Plasma or Other Biological Fluids. This assay employs the competitive inhibition enzyme immunoassay technique. A monoclonal antibody specific to Rat Luteinizing Hormone has been pre-coated onto a microplate. A competitive inhibition reaction is launched between biotin labeled Rat Luteinizing Hormone and unlabeled Rat Luteinizing Hormone (standards or samples) with the pre-coated antibody specific to Rat Luteinizing Hormone. After incubation the unbound conjugate is washed off. Next, Avidin conjugated to Horseradish Peroxidase (HRP) is added to each microplate well and incubated. The amount of bound HRP conjugate is reverse proportional to the concentration of Rat Luteinizing Hormone in the sample. After addition of the substrate solution, the intensity of color developed is reverse proportional to the concentration of Rat Luteinizing Hormone in the sample.

KIT COMPONENTS

Component	Volume
96-well Plate Pre-coated with Anti-Rat Luteinizing Hormone Antibody	8 wells x 12 Strips
Rat Luteinizing Hormone Standard	30 ng x 2
Biotin-Labeled Competitive Inhibitor (100X)	60 µl
Streptavidin-HRP (100X)	120 µl
Standard/Sample Diluent	30 ml
Biotin-Labeled Competitive Inhibitor Diluent	12 ml
Streptavidin-HRP Diluent	12 ml
Wash Buffer (20X)	30 ml
TMB Substrate Solution	12 ml
Stop Solution	12 ml
Plate Adhesive Strips	3 Strips
Technical Manual	1 Manual

STORAGE AND STABILITY

All kit components are stable at 2 to 8 °C. Standard (recombinant protein) should be stored at -20 °C or -80 °C (recommended at -80 °C) after reconstitution. Opened Microplate Wells or reagents may be stored for up to 1 month at 2 to 8 °C. Return unused wells to the pouch containing the desiccant pack, and reseal along the entire edge.

Note: the kit can be used within one year if the whole kit is stored at -20 °C. Avoid repeated freeze-thaw cycles.

MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader capable of measuring absorbance at 450 nm.
2. Adjustable pipettes and pipette tips to deliver 2 µl to 1 ml volumes.
3. Adjustable 1-25 ml pipettes for reagent preparation.
4. 100 ml and 1-liter graduated cylinders.
5. Absorbent paper.
6. Distilled or deionized water.
7. Computer and software for ELISA data analysis.
8. Tubes to prepare standard or sample dilutions.

HEALTH AND SAFETY PRECAUTIONS

1. Reagents provided in this kit may be harmful if ingested, inhaled, or absorbed through the skin. Please carefully review the MSDS for each reagent before conducting the experiment.
2. Stop Solution contains 2 N Sulfuric Acid (H_2SO_4) and is an extremely corrosive agent. Please wear proper eye, hand, and face protection when handling this material. When the experiment is finished, be sure to rinse the plate with copious amounts of running water to dilute the Stop Solution before disposing of the plate.

REAGENT PREPARATION

1. Sample Preparation

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

Cell culture supernates: Remove particulates by centrifugation, assay immediately or aliquot, and store samples at -20°C.

Serum: Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 minutes. Analyze the serum immediately or aliquot and store samples at -20°C.

Plasma: Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 minutes at 1500 X g within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C.

Cell Lysates: Collect cells and rinse cells with PBS. Homogenize and lysate cells thoroughly in lysate solution. Centrifuge cell lysates at approximately 10000 X g for 5 minutes to remove debris. Aliquots of the cell lysates were removed and assayed.

Bone Tissue: Extract demineralized bone samples in 4 M Guanidine-HCl and protease inhibitors. Dissolve the final sample in 2 M Guanidine-HCl.

Tissue Homogenates: The preparation of tissue homogenates will vary depending upon tissue type. Rinse tissue with 1X PBS to remove excess blood, homogenized in 20 mL of 1X PBS, and stored overnight at ≤ -20 °C. After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 5000 x g. The supernate was removed immediately and assayed. Alternatively, aliquot and store samples at ≤ -20 °C.

Note: Some lysis buffers, such as RIPA can not be used. Some components will affect the binding.

Urine: Urinary samples should be cleared by centrifugation and then can be used directly without dilution. Storage at -20°C.

2. Standard Preparation

Reconstitute the lyophilized Rat Luteinizing Hormone Standard by adding 1 ml of Standard/Sample Diluent to make the 30 ng/ml standard stock solution. Allow the solution to sit at room temperature for 5 minutes, then gently vortex to mix completely. Use within one hour of reconstituting. Two tubes of the standard (30 ng per tube) are included in each kit. Use one tube for each experiment.

Perform 2-fold serial dilutions of the top standards to make the standard curve within the range of this assay (0.468 ng/ml - 30 ng/ml) as below. Standard/Sample Dilution Buffer serves as the zero standard (0 ng/ml).

Standard	Add	Into
30 ng/ml		
15 ng/ml	500 µl of the Standard (30 ng/ml)	500 µl of the Standard/Sample Diluent
7.5 ng/ml	500 µl of the Standard (15 ng/ml)	500 µl of the Standard/Sample Diluent
3.75 ng/ml	500 µl of the Standard (7.5 ng/ml)	500 µl of the Standard/Sample Diluent
1.875 ng/ml	500 µl of the Standard (3.75 ng/ml)	500 µl of the Standard/Sample Diluent
0.937 ng/ml	500 µl of the Standard (1.875 ng/ml)	500 µl of the Standard/Sample Diluent
0.469 ng/ml	500 µl of the Standard (0.937 ng/ml)	500 µl of the Standard/Sample Diluent
0 ng/ml	1 ml of the Standard/Sample Diluent	

Note: The standard solutions are best used within 2 hours. The 30 ng/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

3. Biotin-Labeled Competitive Inhibitor Working Solution Preparation

The Biotin-Labeled Competitive Inhibitor should be diluted in 1:100 with the Biotin-Labeled Competitive Inhibitor Diluent and mixed thoroughly. The solution should be prepared no more than 2 hours prior to the experiment.

4. Streptavidin-HRP Working Solution Preparation

The Streptavidin-HRP should be diluted in 1:100 with the Streptavidin-HRP Diluent and mixed thoroughly. The solution should be prepared no more than 1 hour prior to the experiment.

5. Wash Buffer Working Solution Preparation

Pour entire contents (30 ml) of the Wash Buffer Concentrate into a clean 1,000 ml graduated cylinder. Bring the final volume to 600 ml with glass-distilled or deionized water (1:20).

ASSAY PROCEDURE

The Streptavidin-HRP Working Solution and TMB Substrate Solution must be kept warm at 37°C for 30 minutes before use. When diluting samples and reagents, they must be mixed completely and evenly. A standard detection curve should be prepared for each experiment. The user will decide on sample dilution fold by crude estimation of protein amount in samples.

1. Prepare 7 wells for standard, 1 well for blank. Add 50 µl of each standard and samples into appropriate wells. Then add 50 µl of Biotin-Labeled Competitive Inhibitor Working Solution to each well immediately.

2. Cover well and incubate for 60 minutes at room temperature with gentle shaking (using a microplate shaker is recommended).

3. Remove the cover, discard the solution, and wash the plate 3 times with Wash Buffer Working Solution, and each time let Wash Buffer Working Solution stay in the wells for 1 - 2 minutes. Blot the plate onto paper towels or other absorbent material. Do NOT let the wells completely dry at any time.

4. Add 100 µl of Streptavidin-HRP Working Solution into each well and incubate the plate at 37°C for 45 minutes.

5. Wash the plate 5 times with Wash Buffer Working Solution, and each time let the wash buffer stay in the wells for 1 - 2 minutes. Discard the wash buffer and blot the plate onto paper towels or other absorbent material.

6. Add 100 µl of TMB Substrate Solution into each well and incubate the plate at 37°C in the dark for 10 - 20 minutes.

7. Add 100 µl of Stop Solution into each well. The color changes into yellow immediately.

8. Read the O.D. absorbance at 450nm in a microplate reader within 30 minutes after adding the Stop Solution.

For calculation, (the relative O.D.450) = (the O.D.450 of each well) - (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The concentration of the samples can be interpolated from the standard curve.

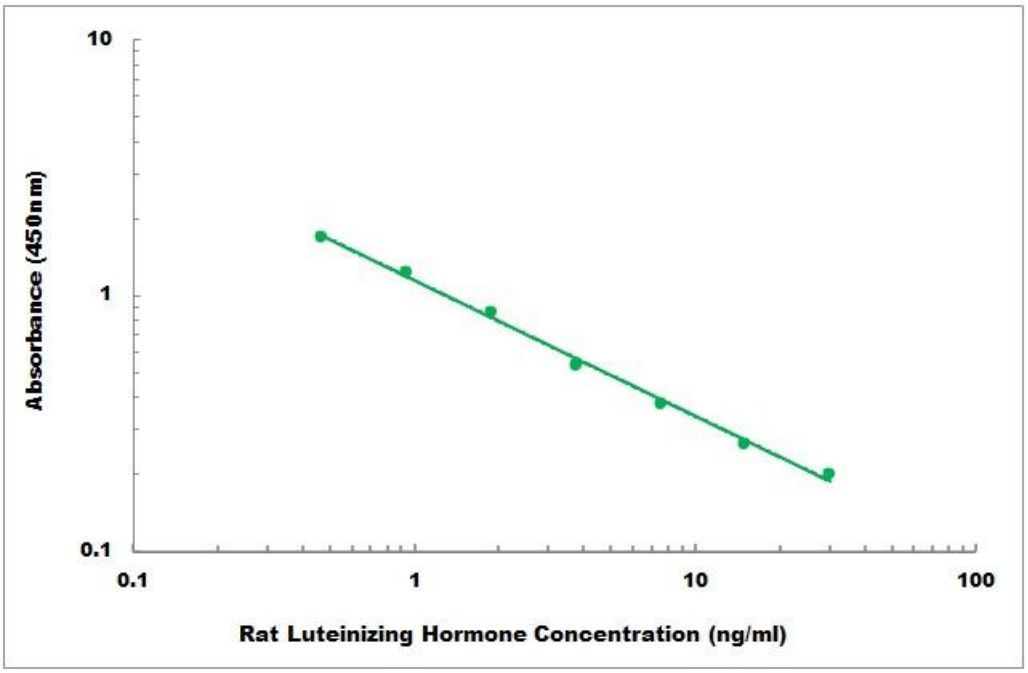
Note: If the samples measured were diluted, multiply the dilution factor by the concentrations from interpolation to obtain the concentration before dilution.

ASSAY PROCEDURE SUMMARY

- Prepare all reagents, samples and standards
- Add 100 µl Standard or Sample
- Wash plate 3 times with Wash Buffer Working Solution
- Add 100 µl Biotin-Labeled Detection Antibody Working Solution
- Wash plate 3 times with Wash Buffer Working Solution
- Add 100 µl Streptavidin-HRP Working Solution
- Wash plate 5 times with Wash Buffer Working Solution
- Add 100 µl TMB Substrate Solution
- Add 100 µl Stop Solution
- Read the plate at 450nm

TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



SENSITIVITY

The minimum detectable dose of Rat Luteinizing Hormone is typically less than 0.14 ng/ml.

SPECIFICITY

Rat Luteinizing Hormone ELISA Kit has high sensitivity and excellent specificity for detection of Rat Luteinizing Hormone.

The detection range is 0.468 ng/ml - 30 ng/ml.

CROSS REACTIVITY

No significant cross-reactivity or interference between Rat Luteinizing Hormone and analogues was observed.

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TROUBLESHOOTING GUIDE

Problem	Possible Cause	Solution
High signal and background in all wells	• Insufficient washing	• Increase number of washes • Increase the time of soaking between in-wash
	• Too much Streptavidin-HRP	• Check dilution, titration
	• Incubation time too long	• Reduce incubation time
	• Development time too long	• Decrease the incubation time before the stop solution is added
No signal	• Reagent added in incorrect order or incorrectly prepared	• Review protocol
	• Standard has gone bad (If there is a signal in the sample wells)	• Check the condition of stored standard
	• Assay was conducted from an incorrect starting point	• Reagents are allowed to come to 20 - 30 °C before performing the assay
Too much signal-whole plate turned uniformly blue	• Insufficient washing-unbound Streptavidin-HRP remaining	• Increase the number of washes carefully
	• Too much Streptavidin-HRP	• Check dilution
	• Plate sealer or reservoir reused, resulting in the presence of residual Streptavidin-HRP	• Use fresh plate sealer and reagent reservoir for each step
Standard curve achieved but poor discrimination between point	• Plate not developed long enough	• Increase substrate solution incubation time
	• Improper calculation of standard curve dilution	• Check dilution, make a new standard curve
No signal when a signal is expected, but the standard curve looks fine	• Sample matrix is masking detection	• More diluted sample Recommended
Samples are reading too high, but the standard curve is fine	• Samples contain protein levels above the assay range	• Dilute samples and run Again

Edge effect	• Uneven temperature around the work surface	• Avoid incubating plates in areas where environmental conditions vary • Use plate sealer